# EnzyFluo<sup>™</sup> Uricase Assay Kit (EURIC-100)

**Quantitative Fluorimetric Uricase Activity Determination** 

## DESCRIPTION

URICASE or URATE OXIDASE (E.C. 1.7.3.3) initiates the oxidation of uric acid, eventually yielding allantoin. Uricase activity is found throughout plants, animals, and bacteria, but it is not found in hominids. As a result, humans develop gout when the levels of uric acid become too high. BioAssay Systems' method provides a simple and high-throughput assay for measuring uricase enzyme activity. In this assay, uric acid is enzymatically converted to allantoin, releasing H<sub>2</sub>O<sub>2</sub>. The resulting H<sub>2</sub>O<sub>2</sub> reacts with a specific dye to form a pinkcolored product. The change in fluorescence intensity at 530/585 nm is directly proportional to the uricase activity in the sample.

## **KEY FEATURES**

Sensitive. Use 10  $\mu L$  samples. Linear detection range: fluorimetric assay 0.3 - 100 U/L Uricase.

Fast. Run time is 10 minutes for rapid results.

**Convenient.** Room temperature "mix-and-read" procedure can be readily automated for high-throughput assay of thousands of samples per day.

## **APPLICATIONS**

**Assays:** Uricase activity in biological samples such as cell culture, non-human biofluids, cell extracts, and plant tissue extracts.

## **KIT CONTENTS**

 Assay Buffer: 5 mL
 Dye Reagent: 120 μL

 Substrate: 5 mL 1mM Uric Acid
 HRP Enzyme: 120 μL

 Uricase Control: 20 μL
 Standard: 100 μL 3% H<sub>2</sub>O<sub>2</sub>

**Storage conditions**. The kit is shipped on ice. Store all components at -20°C. Shelf life of six months after receipt.

**Precautions:** Reagents are for research use only. Normal precautions for laboratory reagents should be exercised while using the reagents. Please refer to Material Safety Data Sheet for detailed information.

## FLUORIMETRIC ASSAY

Liquid samples such as serum and plasma can be assayed directly. Tissue and cell lysates need to be clear and debris-free.

Note: SH-containing reagents (e.g.  $\beta$ -mercaptoethanol, dithiothreitol, > 5  $\mu$ M), sodium azide, EDTA, and sodium dodecyl sulfate are known to interfere in this assay and should be avoided in sample preparation.

- 1. Equilibrate all components to room temperature. Briefly centrifuge the tubes before opening. Keep thawed tubes on ice during assay.
- 2.  $H_2O_2$  Standard Curve. Mix 10 µL 3%  $H_2O_2$  and 870 µL dH<sub>2</sub>O (final 10 mM) then mix 10 µL of the 10 mM  $H_2O_2$  with 990 µL dH<sub>2</sub>O to yield 100 µM  $H_2O_2$ . Prepare standards as shown in the Table below.

No	100 µM H <sub>2</sub> O <sub>2</sub> + H <sub>2</sub> O	Vol (µL)	H <sub>2</sub> O <sub>2</sub> (μM)
1	100 μL + 0 μL	100	100
2	60 μL + 40 μL	100	60
3	30 μL +   70 μL	100	30
4	0 μL + 100 μL	100	0

Transfer 10  $\mu L$  standards and samples into separate wells of a black flat-bottom 96-well plate. Dilute the Uricase Control 1:100 in Assay Buffer then transfer 10  $\mu L$  of the enzyme to the Control well.

- Fluorescence reaction. Prepare enough Working Reagent by mixing, for each well, 50 μL Assay Buffer, 50 μL 1 mM Uric Acid Substrate, 1 μL HRP enzyme and 1 μL Dye Reagent. Add 90 μL Working Reagent to each well.
- 4. Read fluorescence intensity at  $\lambda_{ex}$  = 530 nm and  $\lambda_{em}$  = 585 nm in kinetic mode for 10 minutes.

*Note*: If the observed uricase activity is higher than 100 U/L, dilute sample in Assay Buffer and repeat the assay. Multiply result by the dilution factor n.

## CALCULATION

Subtract blank  $F_{10}$  (water, #4) from all standard  $F_{10}$  values and plot the  $\Delta F$  against standard concentrations. Determine the slope using linear regression. Calculate the  $\Delta F_{Sample}$  of all samples by subtracting  $F_0$  from  $F_{10}$  for each sample. Do the same for the blank (water, standard #4) to get  $\Delta F_{Blank}.$  Calculate the activity using the equation below:

Uricase Activity = 
$$\frac{\Delta F_{\text{SAMPLE}} - \Delta F_{\text{BLANK}}}{\text{Slope} (\mu M^{-1}) \cdot t} \times \frac{\text{Reaction Vol.}}{\text{Sample Vol.}} \times n (U/L)$$
  
=  $\frac{\Delta F_{\text{SAMPLE}} - \Delta F_{\text{BLANK}}}{\text{Slope} (\mu M^{-1})} \times n (U/L)$ 

Where  $\Delta F_{\text{Sample}}$  and  $\Delta F_{\text{Blank}}$  are the changes in fluorescence values of the sample and blank, respectively. *Slope* is the slope of the H<sub>2</sub>O<sub>2</sub> standard curve, *t* is the incubation time (10 minutes), and *n* is the dilution factor. Note: in the standard protocol, the reaction volume/sample volume value cancels out the time value.

Unit definition: 1 U Uricase will generate 1  $\mu mol$  of  $H_2O_2$  from Uric Acid per minute at 25 °C at pH 7.5.

#### MATERIALS REQUIRED, BUT NOT PROVIDED

Uricase-containing samples, pipetting devices, centrifuge tubes, black flat-bottom uncoated 96-well plates, and fluorescence plate reader.



*Left*: 96-Well Fluorimetric Assay. ■ 100U/L Uricase, ♦ 60 U/L Uricase, ● Rat Liver Microsomes, ▼ Pea Roots. *Right*: Uricase Specific Activity in rat liver cytosol (#1), rat liver S9 fraction (#2), rat liver microsomes (#3), pea roots (#4) and pea shoots (#5)

## LITERATURE

1. Kratzer JT et al (2014) Evolutionary history and metabolic insights of ancient mammalian uricases. Proc Natl Acad Sci USA 111(10):3763-8.

2. Maiuolo J et al (2016) Regulation of uric acid metabolism and excretion. Int J Cardiol. 213:8-14

3. Roman YM (2023) The role of uric acid in human health: Insights from the Uricase gene. J Pers Med. 13(9):1409

