

## EnzyFluo™ Farnesyltransferase Inhibitor Screening Kit (EIFT-400)

### Fluorimetric Inhibitor Screening Assay for Farnesyltransferase

#### DESCRIPTION

**FARNESYLTRANSFERASE (FTase, EC 2.5.1.58)** catalyzes the transfer of a farnesyl group from farnesyl pyrophosphate to the cysteine residue of the C-terminus of target proteins. When not properly regulated, farnesylated proteins, including the Ras superfamily of small GTPases, can lead to developmental disorders and cancer. Simple, direct and high-throughput inhibitor screening assays find wide applications for oncology research. BioAssay Systems' EIFT-400 assay kit provides a convenient fluorimetric method to screen for potential FTase inhibitors. In this assay, FTase reacts with farnesyl pyrophosphate and a dansyl-peptide substrate with measurable fluorescence at  $\lambda_{em/ex} = 340/550$  nm. Inhibition is determined by the decrease in fluorescence.

#### KEY FEATURES

**Safe.** Non-radioactive assay.

**Homogeneous and convenient.** "Mix-incubate-measure" type assay. No wash and reagent transfer steps are involved.

**High-throughput.** A Z'-factor of 0.8 and higher is routinely observed in a 384-well format. Can be readily automated to assay thousands of samples per day.

#### APPLICATIONS

For high-throughput screening of FTase inhibitors and evaluation of drug modulators.

#### KIT CONTENTS (400 TESTS IN 384-WELL PLATES)

<b>Assay Buffer:</b>	12 mL	<b>Substrate:</b>	200 $\mu$ L
<b>150mM DTT:</b>	400 $\mu$ L	<b>10mM Lonafarnib:</b>	50 $\mu$ L

**Storage conditions.** The kit is shipped on ice. Store all components at  $-20^{\circ}\text{C}$ . Shelf life: 6 months.

**Precautions:** Reagents are for research use only. Normal precautions for laboratory reagents should be exercised while using the reagents. Please refer to Material Safety Data Sheet for detailed information.

#### PROCEDURES

This assay is based on an enzyme-catalyzed kinetic reaction. To ensure identical incubation times, the addition of Working Reagent should be quick and mixing should be brief but thorough. Use of a multichannel pipettor is recommended. *Note: FTase enzyme is not included in the kit.*

**Reagent Preparation:** Use black, flat-bottom 384-well plates. Prior to assay, equilibrate all components to room temperature and briefly centrifuge tubes before opening. The Working Reagent (WR) should be prepared fresh for each assay run.

**Enzyme Preparation:** Enzyme should be prepared in buffer and used fresh. The following protocol is optimized for recombinant rat FTase enzyme (Jena Bioscience, Cat #PR-102). If using a different enzyme, we recommend that you experimentally determine the optimal amount of enzyme to use per well.

**Test Compound Preparation:** Dissolve test compounds in a solvent of choice, e.g. DMSO. It is prudent to first test the tolerance of the enzyme with the solvent of choice. In the example below, rat FTase (Jena Bioscience, Cat #PR-102) was found to tolerate up to 2v% DMSO. Lonafarnib is included as an  $\text{IC}_{50}$  control.

1. **Enzyme and Controls.** Transfer 5  $\mu$ L of 37.2  $\mu$ g/mL FTase (7.4  $\mu$ g/mL final concentration) into separate wells of a black 384-well plate.

Reserve at least one well for no inhibitor (Control) and one with no enzyme (Blank). Add 5  $\mu$ L of 37.2  $\mu$ g/mL FTase and 5  $\mu$ L of Assay Buffer to the Control and Blank wells, respectively. We recommend running at least duplicate reactions.

2. **Add Test Compounds.** To the enzyme wells, add 5  $\mu$ L of the test compounds.

To the Control and Blank wells, add 5  $\mu$ L of the solvent that the test compounds are dissolved in. For example, if the test compounds are dissolved in 1% DMSO, add 5  $\mu$ L of 1% DMSO to these wells.

Mix *immediately* and incubate for 10 min at RT for test compounds to interact with FTase.

3. Prepare enough WR for all wells by mixing for each well, 0.5  $\mu$ L Substrate, 20  $\mu$ L Assay Buffer and 1  $\mu$ L DTT. Transfer 15  $\mu$ L WR to all wells. *Immediately* tap the plate to mix and incubate at RT.

4. Read fluorescence intensity at 60 min at  $\lambda_{ex/em} = 340/550\text{nm}$ .

#### CALCULATION

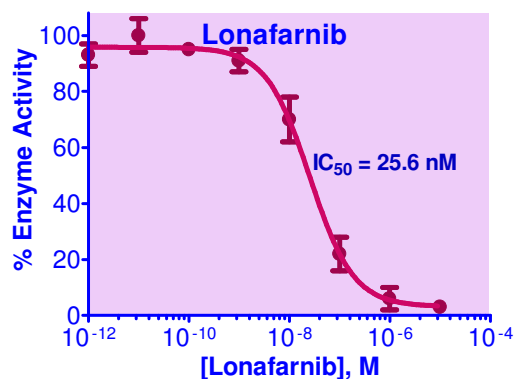
FTase activity in the presence of test compounds is calculated as follows,

$$\% \text{ Activity} = \frac{\text{RFU}_{\text{Test Cpd}} - \text{RFU}_{\text{Blank}}}{\text{RFU}_{\text{Control}} - \text{RFU}_{\text{Blank}}} \times 100\%$$

where the  $\text{RFU}_{\text{Test Cpd}}$ ,  $\text{RFU}_{\text{Control}}$ , and  $\text{RFU}_{\text{Blank}}$  are the fluorescence values of the test compound, no inhibitor control, and no enzyme blank at 60 min.

#### MATERIALS REQUIRED, BUT NOT PROVIDED

Pipetting devices and accessories (e.g. multi-channel pipettor), black, flat-bottom 384-well plates (e.g. Corning™ 3573 cat# 09-761-86), centrifuge tubes and plate reader. FTase enzyme is not included.



Recombinant Rat FTase was incubated with various concentrations of Lonafarnib. Each concentration of inhibitor contained 1v% DMSO. The  $\text{IC}_{50}$  of Lonafarnib with 7.4  $\mu$ g/mL FTase enzyme was determined to be 25.6 nM.

#### LITERATURE

- [1]. Appels, Natalie et al (2005). Development of farnesyl transferase inhibitors: a review. *Oncologist*: vol. 10(8) 565-78.
- [2]. Long, Stephen et al (2001). The crystal structure of human protein farnesyltransferase reveals the basis for inhibition by CaaX tetrapeptides and their mimetics: *PNAS*: vol. 98(23) 12948-12953.
- [3]. Wang, Jingyuan et al (2017). New tricks for human farnesyltransferase inhibitor: cancer and beyond. *MedChemComm*: vol. 8(5) 841-854.

