

## EnzyChrom™ D-Lactate Assay Kit (EDLC-100)

### Colorimetric Determination of D-Lactate at 565 nm

#### DESCRIPTION

Lactate is generated by lactate dehydrogenase (LDH) under hypoxic or anaerobic conditions. Monitoring lactate levels is, therefore, a good indicator of the balance between tissue oxygen demand and utilization and is useful when studying cellular and animal physiology. D-lactate is produced in only minor quantities in animals and measuring for D-lactate in animal samples is a means to determine the presence of bacterial infection.

Simple, direct and automation-ready procedures for measuring lactate concentration are very desirable. BioAssay Systems' EnzyChrom™ lactate assay kit is based on lactate dehydrogenase catalyzed oxidation of lactate, in which the formed NADH reduces a formazan (MTT) Reagent. The intensity of the product color, measured at 565 nm, is proportionate to the lactate concentration in the sample.

#### APPLICATIONS

**Direct Assays:** D-lactate in serum, plasma, and cell media samples.

#### KEY FEATURES

**Sensitive and accurate.** Detection limit of 0.05 mM and linearity up to 2 mM D-lactate in 96-well plate assay. *For cell culture samples containing phenol red:* detection limit of 0.1 mM and linearity up to 1 mM D-lactate in 96-well plate assay.

**Convenient.** The procedure involves adding a single working reagent, and reading the optical density at time zero and at 20 min. Room temperature assay. No 37°C heater is needed.

**High-throughput.** Can be readily automated as a high-throughput 96-well plate assay for thousands of samples per day.

#### KIT CONTENTS (100 tests in 96-well plates)

<b>Assay Buffer:</b> 10 mL	<b>NAD Solution:</b> 1 mL
<b>Enzyme A:</b> 120 µL	<b>MTT Solution:</b> 1.5 mL
<b>Enzyme B:</b> 120 µL	<b>Standard:</b> 1.0 mL 20 mM D-lactate

**Storage conditions.** The kit is shipped on ice. Store all reagents at -20°C. Shelf life: 6 months after receipt.

**Precautions:** reagents are for research use only. Normal precautions for laboratory reagents should be exercised while using the reagents. Please refer to Material Safety Data Sheet for detailed information.

#### PROCEDURES

**Important:** this assay is based on an enzyme-catalyzed kinetic reaction. Addition of Working Reagent should be quick and mixing should be brief but thorough. Use of a multi-channel pipettor is recommended.

- Standard Curve.** Prepare 1000 µL 2.0 mM D-lactate Premix by mixing 100 µL 20 mM Standard and 900 µL distilled water. *For cell culture samples containing phenol red,* prepare 1000 µL 1.0 mM lactate Premix by mixing 50 µL 20 mM Standard and 950 µL culture medium *without serum.* Dilute standard as follows. Transfer 20 µL standards into wells of a clear bottom 96-well plate.

No	Premix + H <sub>2</sub> O or Medium	Vol (µL)	D-lactate (mM)
1	100µL + 0µL	100	2.0 or 1.0
2	80µL + 20µL	100	1.6 or 0.8
3	60µL + 40µL	100	1.2 or 0.6
4	40µL + 60µL	100	0.8 or 0.4
5	30µL + 70µL	100	0.6 or 0.3
6	20µL + 80µL	100	0.4 or 0.2
7	10µL + 90µL	100	0.2 or 0.1
8	0µL + 100µL	100	0

**Samples.** Add 20 µL sample per well in separate wells. For samples with potential endogenous enzyme activity (i.e. serum, plasma,

tissue extracts), two reactions should be run: one with added Enzyme A and a No Enzyme A control. Serum and Plasma should be diluted at least 2× with dH<sub>2</sub>O prior to assay.

- Reagent Preparation.** Spin the Enzyme tubes briefly before pipetting. For each Sample and Standard well, prepare Working Reagent by mixing 60 µL Assay Buffer, 1 µL Enzyme A, 1 µL Enzyme B, 10 µL NAD and 14 µL MTT. Fresh reconstitution is recommended. For the No Enzyme A control, the Working Reagent includes 60 µL Assay Buffer, 1 µL Enzyme B, 10 µL NAD and 14 µL MTT.
- Reaction.** Add 80 µL Working Reagent per reaction well quickly. Tap plate to mix briefly and thoroughly.
- Read optical density (OD<sub>0</sub>)** for time "zero" at 565 nm (520-600nm) and OD<sub>20</sub> after a 20-min incubation at room temperature.
- Calculation.** Subtract OD<sub>0</sub> from OD<sub>20</sub> for the standard and sample wells. Use the ΔOD values to determine the sample D-lactate concentration from the standard curve. For samples requiring a No Enzyme A control, subtract the ΔOD<sub>NoEnz</sub> value from the ΔOD<sub>Sample</sub> and use this ΔΔOD value to determine the sample D-lactate concentration from the standard curve.

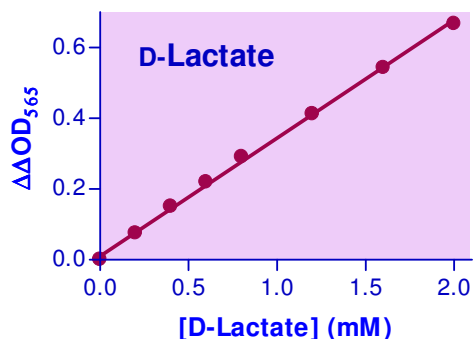
Note: if the sample OD value is higher than OD for 2 mM D-lactate standard, dilute sample in water and repeat the assay. Multiply the results by the dilution factor.

#### MATERIALS REQUIRED, BUT NOT PROVIDED

Pipeting (multi-channel) devices. Clear-bottom 96-well plates (e.g. Corning Costar) and plate reader.

#### GENERAL CONSIDERATIONS

The following substances interfere and should be avoided in sample preparation: EDTA (>0.5 mM), ascorbic acid, SDS (>0.2%), sodium azide, NP-40 (>1%) and Tween-20 (>1%).



Standard Curve in 96-well plate assay in water.

#### PUBLICATIONS

- Lee, JH et al (2020). Effect of cysteine on methylglyoxal-induced renal damage in mesangial cells. *Cells*, 9(1).
- Aranda-Diaz, A et al (2020). Bacterial interspecies interactions modulate pH-mediated antibiotic tolerance. *ELife*, 9.
- Marcu, R et al (2014). Mitochondrial Matrix Ca<sup>2+</sup> Accumulation Regulates Cytosolic NAD/NADH Metabolism, Protein Acetylation, and Sirtuin Expression. *Mol Cell Biol*. 34(15):2890-902.

